

Abstract

Metabotropic glutamate receptor 7 (mGlu₇) is a dimeric, group III metabotropic glutamate (mGlu) receptor. It is a G protein-coupled receptor that acts to modulate neurotransmission across many brain structures. mGlu₇ is most highly expressed presynaptically in neurons and is widely distributed in the central nervous system (CNS) and body. Mutations, deletions, or decreases in mGlu₇ result in symptoms and phenotypes of neurodevelopmental disorders in humans and mice¹, including Rett syndrome. We are focused on developing new ligands that interact with mGlu₇ to understand the therapeutic potential of the receptor in various CNS disorders.

We are using two *in vitro* molecular pharmacology assays to test newly developed compounds for activity at mGlu₇. However, these and other *in vitro* studies show that the orthosteric ligand, glutamate, has low affinity for mGlu₇, which limits its ability to be routinely used for drug screening. An alternative agonist, L-AP4, is a synthetic compound that, while not produced naturally *in vivo*, activates mGlu₇ with higher affinity than glutamate. Our preliminary research indicates, however, that there are differences in the profiles of positive allosteric modulators (PAMs) when assessed using glutamate versus L-AP4. This difference creates a challenge for the development of mGlu₇-selective PAMs, and the focus of our current studies is to understand the difference between these two agonists in activating the mGlu₇ receptor.

Previous literature shows that some mGlu receptors demonstrate partial activity when only one agonist binding site is occupied and full activity when binding to both agonist binding sites occurs within an mGlu receptor dimer². Therefore, we hypothesize that glutamate and L-AP4 activate mGlu₇ either via a different number of agonist binding sites or a different number of effector (G-protein) binding sites. Our current studies are designed to differentiate between these two possibilities.

mGlu₇ responds differently to Glutamate and L-AP4

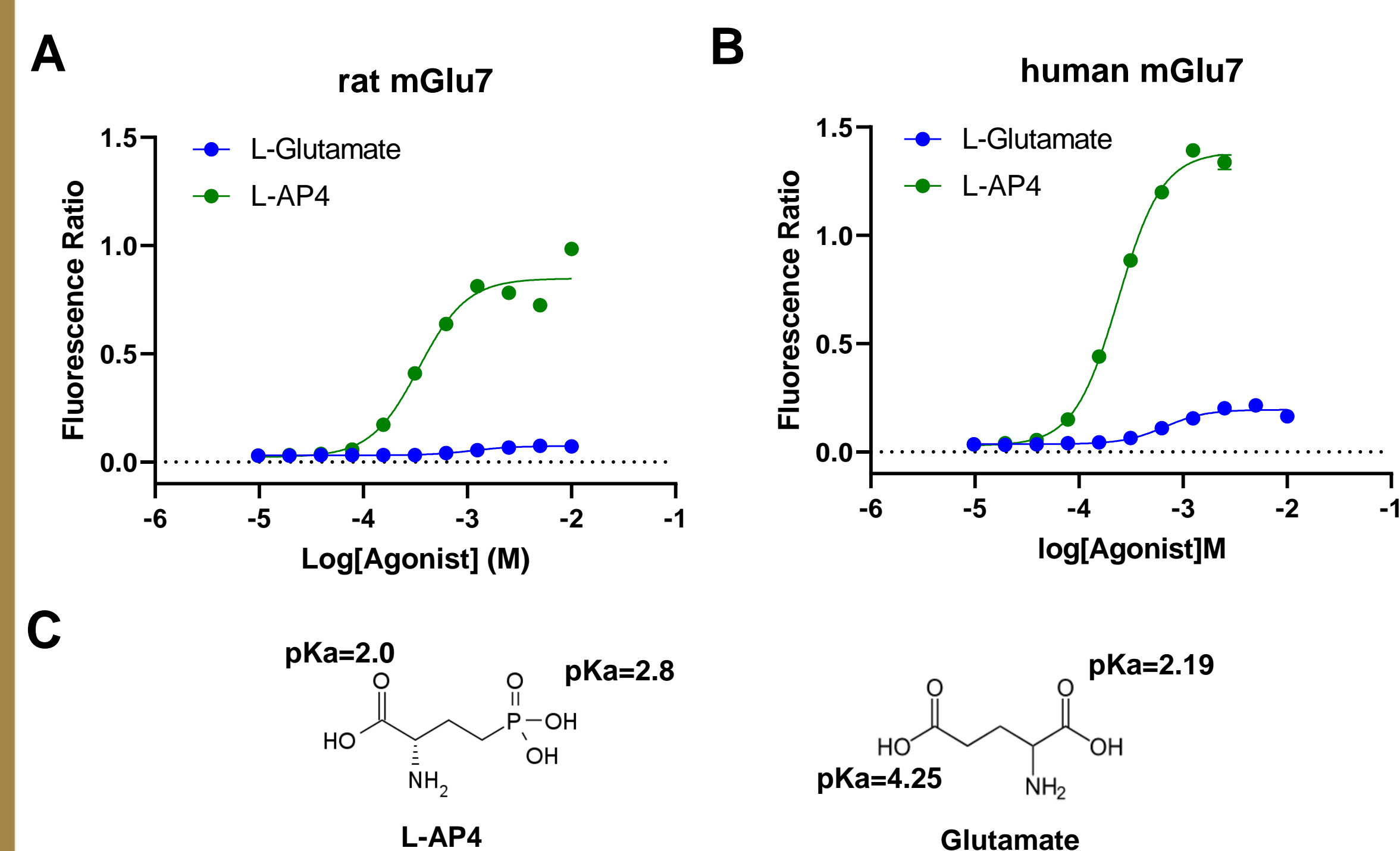


Figure 1. Low affinity and functional potency of glutamate at mGlu₇ necessitates an alternative agonist for functional drug screening. A. Concentration-response curve (CRC) of wild-type (WT) rat mGlu₇ with different agonists in HEK Gα15 cells as measured using a calcium mobilization assay. B. CRC of different agonists at WT human mGlu₇. C. Chemical structure of L-AP4 and Glutamate

References

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- Margeta-Mitrovic, M., Jan, Y. N. & Jan, L. Y. Function of GB1 and GB2 subunits in G protein coupling of GABAB receptors. *Proc. Natl. Acad. Sci. U. S. A.* **98**, 14649–14654 (2001).

Heterodimer restriction of mGlu₇ subunits

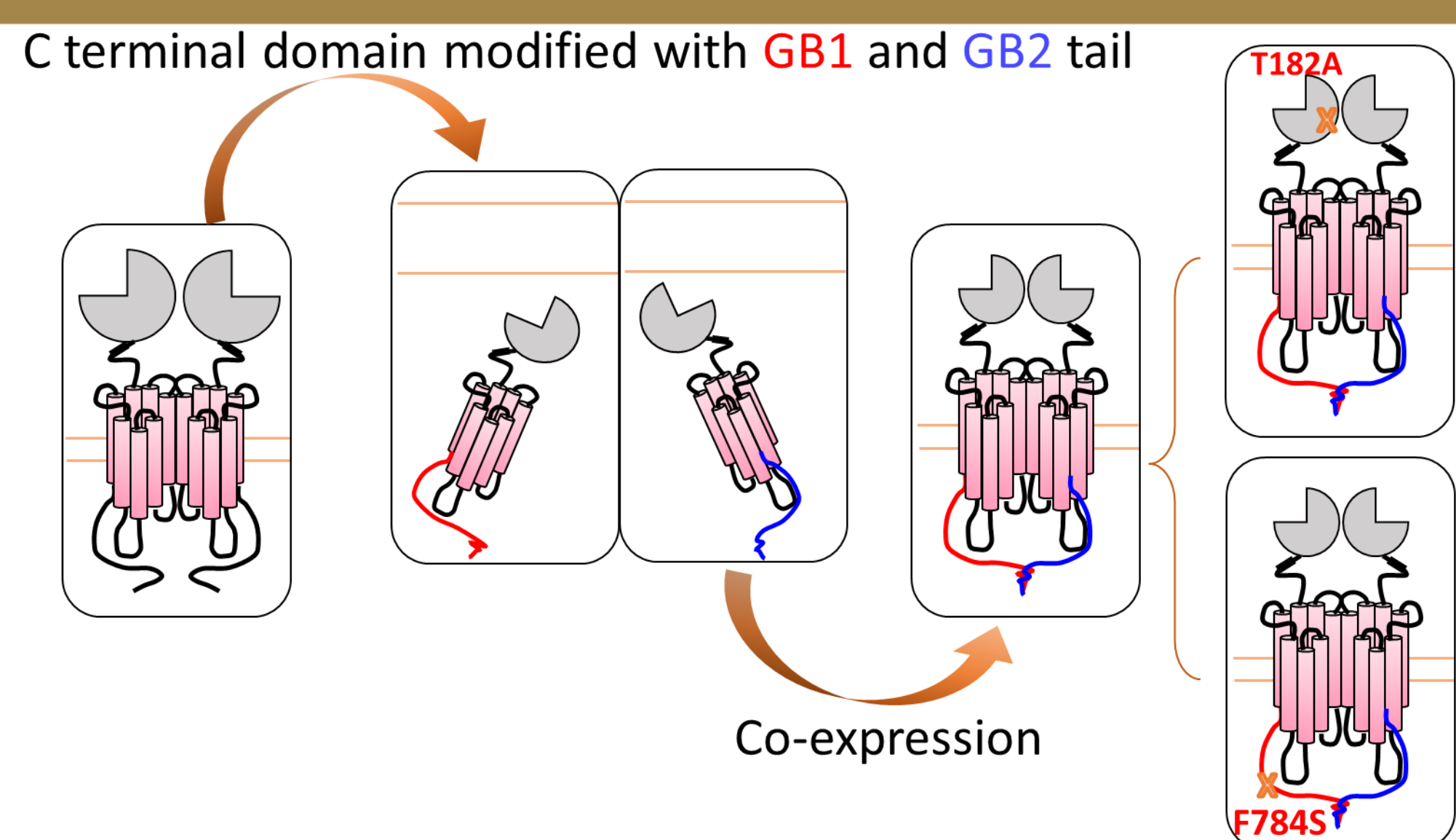


Figure 2. mGlu₇ constructs were designed for expression with hemagglutinin (HA) tags on the N terminus. The C-terminal domains of the receptors were modified to contain tails derived from the two GABA_B receptor subunits, GB1 and GB2; this strategy permits only heterodimers with this two subunit combination to traffic to the cell surface³. Individual mGlu₇ receptor subunits were then mutated at either the agonist binding site (T182A) or the G-protein activation site (F784S).

Modified mGlu₇ dimers are expressed on the cell surface

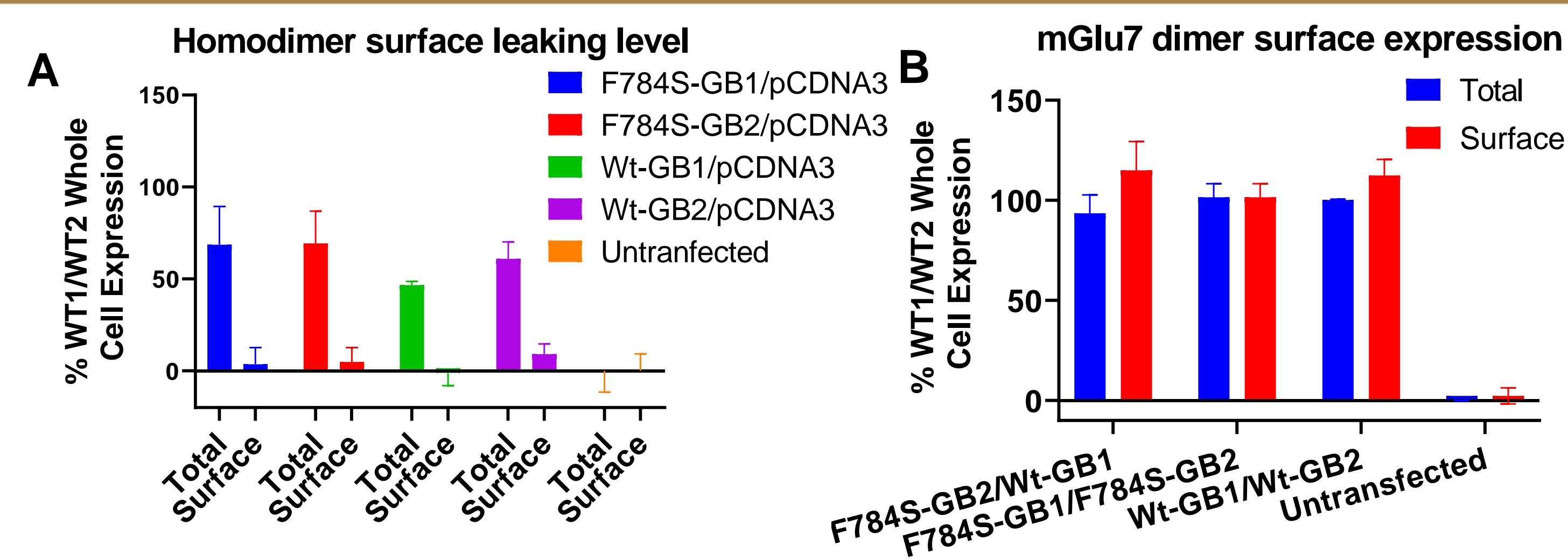


Figure 3. Evaluation of total and surface expression of mGlu₇ by on cell ELISA. A. When expressed alone, mGlu₇ receptors with either a GABA_{B1} or GABA_{B2} tails are expressed in cells but do not travel to the surface by themselves (N=1). B. When GB1 and GB2 tails are expressed together, dimers traffic efficiently to the cell surface (N=3).

Glutamate and L-AP4 activate G-protein binding sites differently

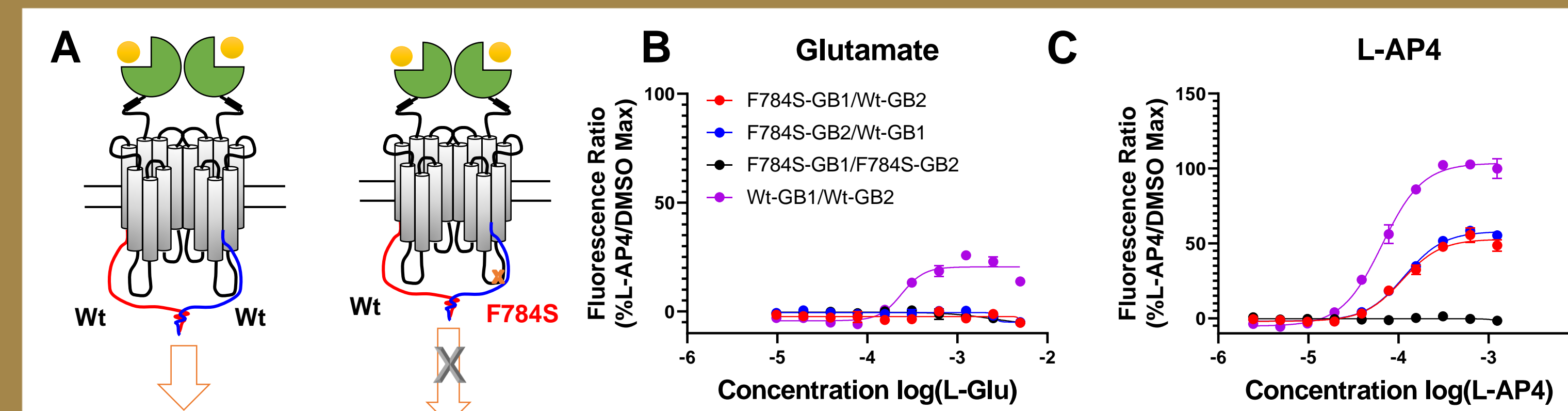


Figure 4. A. Schematic of mGlu₇ dimer activation both as a wild-type (WT) receptor tagged with GB1 (red) and GB2 (blue) tails and with an F784S mutation to disrupt G protein coupling in the GB2 subunit. Activation of various mGlu₇ dimers with glutamate (B) or L-AP4 (C). While L-AP4 is able to activate a receptor in which one G protein binding site has been eliminate, glutamate can not (N=1).

PAMs differentially affect receptor signaling with distinct agonists

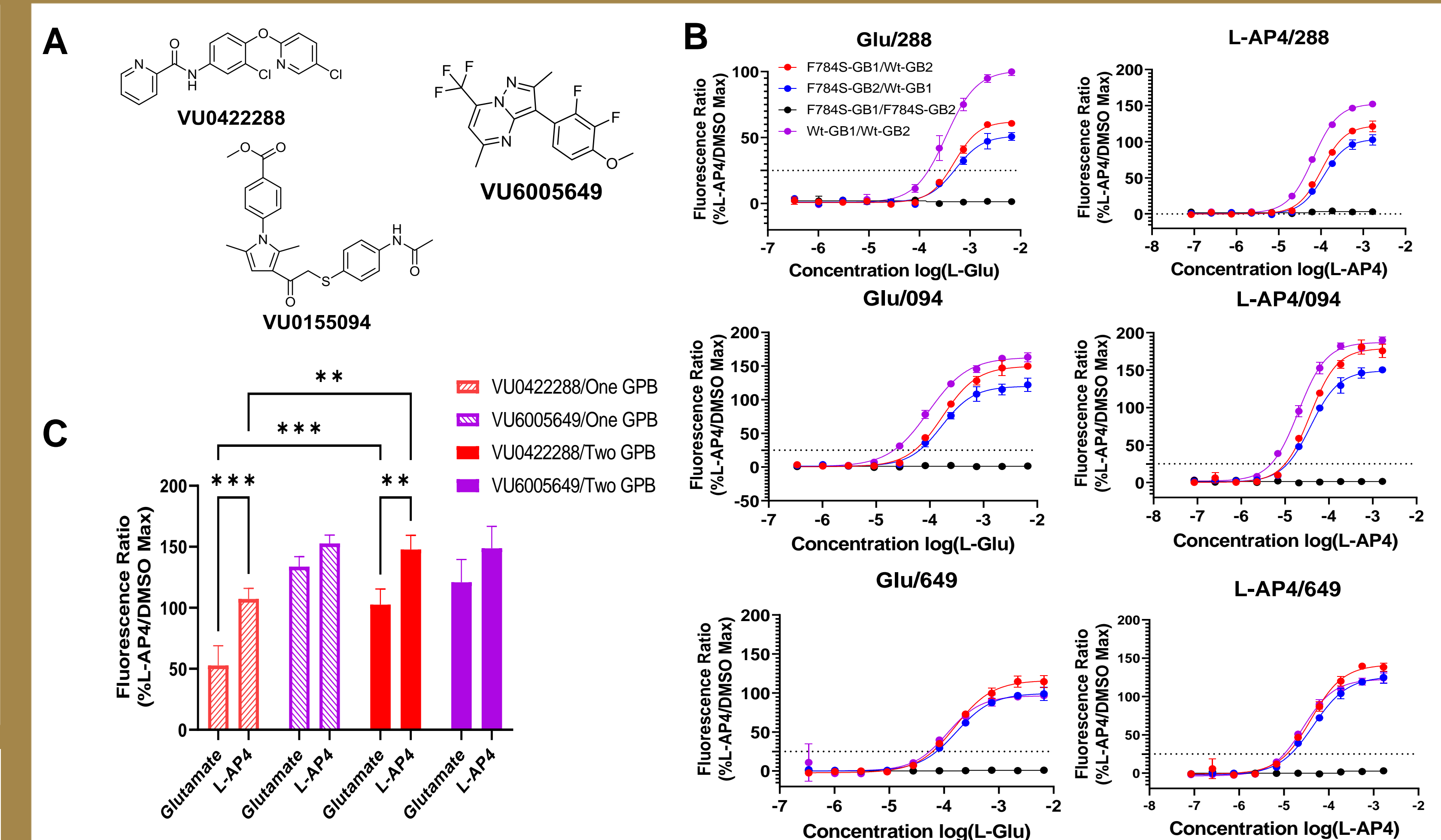


Figure 5. PAMs differentially affect G-protein activation. A. Structure of positive allosteric modulators (PAMs) used in these studies. VU288 and VU094 are group III mGlu PAMs and VU649 is mGlu_{7/8} PAM. B. CRC of different mGlu₇ dimer responses to glutamate and L-AP4. C. The maximal responses of VU288 and VU649 from Ca assays with different agonists exhibit significant differences. N=4, 2-way ANOVA with Turkey post-test, **p<0.01, ***p<0.001, ****p<0.0001

Table 1 LogEC50 from Ca assays with different agonists N=3

Dimers	DMSO	VU0422288	VU0155094	VU6005649
F784S-GB1/Wt-GB2	N/A	-3.61±0.14	-3.35±0.08	-4.01±0.05
F784S-GB2/Wt-GB1	N/A	-3.62±0.10	-3.34±0.10	-4.00±0.06
F784S-GB1/F784S-GB2	N/A	N/A	N/A	N/A
Wt-GB1/Wt-GB2	-3.14±0.09	-3.81±0.09	-3.54±0.06	-4.24±0.09
Agonists	Glutamate	L-AP4	Glutamate	L-AP4

Discussion and future plans

- Different agonists activate mGlu₇ distinctly based on the number of intact G-protein activation sites.
- Glutamate activates mGlu₇ only when both G-protein binding sites are functional, while L-AP4 activates the receptor partially with one G-protein binding site.
- The PAMs also show differential effects on signaling, with some PAMs requiring two G-protein binding sites to induce full activation while others only require one.

Studies with glutamate binding-site mutations (T182A) are currently underway, as well as studies regarding how each mGlu₇ protomer is activated within heterodimeric receptors.

Acknowledgements

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